

p-Hydroxyphenacyl Phototriggers: The Reactive Excited State of Phosphate Photorelease

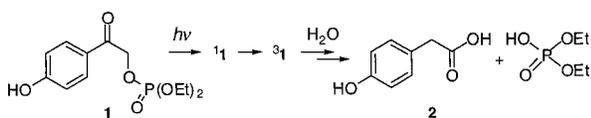
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Fast release of biological stimulants is needed to monitor physiological response in real time.¹ *p*-Hydroxyphenacyl (*p*HP) phototriggers meet this requirement. Moreover, the side products are biologically compatible and transparent to excitation wavelengths >300 nm.² Knowledge of the release rate is essential to biophysical applications. Givens et al. identified a short-lived triplet state as the reactive excited state of *p*HP phosphates and carboxylates on the basis of quenching experiments.² In a recent study of *p*HP esters Zhang et al.^{3a} questioned that evidence and favored a singlet mechanism. Here, we prove that the reactive excited state of *p*HP diethyl phosphate (**1**) is a very short-lived triplet state and we provide direct evidence for adiabatic tautomerization of *p*-hydroxyacetophenone and its derivatives in the triplet state.



p-Hydroxyphenylacetic acid (**2**) is the only product detected by GC–MS after irradiation of **1** in wet acetonitrile (AN, H₂O ≥ 5% by vol). Numerous products of higher molecular weight are formed in dry AN. Relative quantum yields of the photoreaction in different media were determined by the permanent bleaching of the absorption by **1** at 270 nm (pulsed laser excitation at 308 nm). Water (5%) accelerated the reaction about 5-fold, but saturation with air or addition of 0.5 mM piperylene slowed the conversion about 4-fold in dry AN. The absolute quantum yield in wet AN (5%) was determined as 0.94 (steady-state 313-nm irradiation, azobenzene actinometry); it was reduced to 0.37 by addition of 10 mM piperylene. Still higher quencher concentrations were required to reduce the quantum yield in solutions containing more water.^{2c,3a}

Identification of the transients produced by nanosecond laser flash photolysis (LFP)⁴ of **1** was aided by referring to *p*-hydroxyacetophenone (**3**) and *p*-methoxyacetophenone (**4**) as model compounds. Strong absorption was formed by LFP of **4** (1 × 10⁻⁴ M) in aqueous AN (50% H₂O, λ_{max} ≈ 395 nm) or AN (385 nm). Admission of air reduced the lifetime of this transient

from ~30 μs to 80 ns in AN, and it was identified as the triplet state of **4**, **3**₄, by energy transfer to naphthalene (*E*_T = 61 kcal mol⁻¹),⁵ *k*_{et} ≈ 1 × 10¹⁰ M⁻¹ s⁻¹. Pump–probe spectroscopy of **4** in AN (248-nm excitation, 1 mJ per pulse, 0.8 ps pulse length) showed that intersystem crossing (ISC), **1**₄ → **3**₄, is very fast, *k*_{ISC} = (3.1 ± 0.2) × 10¹¹ s⁻¹.

Replacement of the methoxy group by an ionizable hydroxy group introduces new functionality (Scheme 1). The ionization constant of **3** is p*K*_a(**3**) = 7.9 ± 0.1.⁶ The equilibrium constant for the isomerization of **3** to its quinonoid enol tautomer **3**[′], p*K*_E = 16.4, was calculated using density functional theory (DFT).⁷ A thermodynamic cycle then defines the acidity of enol **3**[′] to be very high, p*K*_a(**3**[′]) = -8.5.

Formation of **3**³ is equally fast in aqueous AN solvent mixtures (*k*_{ISC} = 2.7 × 10¹¹ s⁻¹, λ_{max} = 370 in dry AN, 395 nm with 50% H₂O). Energy transfer from **3**³ to naphthalene was again observed.⁵ The transient is acidic,⁸ but does not ionize in dry AN. Adiabatic ionization, **3**³ → **3**³⁻ + H⁺, takes place in aqueous AN (50% H₂O), *k*_{ion} ≈ 9 × 10⁶ s⁻¹, and is too fast for detection by LFP in water, *k*_{ion} > 3 × 10⁷ s⁻¹. The spectra of **3**³ and **3**³⁻ appear to be quite similar; the absorption maximum of **3**³⁻ is shifted to slightly longer wavelength (405 nm) and the weaker band in the visible, λ_{max} ≈ 500 nm, is more pronounced. In a second process, *k* = 1.9 × 10⁶ s⁻¹, the intensity of the 405-nm band is reduced, and a new strong band appears at 350 nm.^{3b} Excitation of the anion **3**⁻ in aqueous NaOH (0.1 M) at 308 nm gave only the 405 and 500 nm bands, which shows that these are due to the triplet state of the anion, **3**³⁻.

Apparently, **3**³⁻ is re-protonated to form the 350-nm transient in acidic solutions. The first-order rate coefficient for re-protonation in dilute aqueous acid ([HClO₄] = 3–10 × 10⁻⁴ M) was proportional to acid concentration, *k*_{H⁺} ≈ 4 × 10¹⁰ M⁻¹ s⁻¹. Spectrographic traces of the equilibrated triplet–triplet absorptions (delay relative to excitation 200 ns) formed by excitation of **3** in aqueous buffer solutions at pH 7.1 (phosphate, *I* = 0.01 M), 5.32, 4.66, 4.38, 4.04 (acetate, *I* = 0.01 M), and 2.0 (HClO₄, 10⁻² M) are shown in Figure 1. Factor analysis indicated that two components sufficed to reproduce all spectra within experimental accuracy. Fitting of a titration function to the loading coefficients gave p*K*_a = 4.6 ± 0.2.

We attribute the protonated species observed at low pH (λ_{max} ≈ 350 nm) to the quinonoid enol triplet, **3**^{3′}. Clearly, **3**^{3′} is an excited triplet—it is in equilibrium with **3**³⁻ and the lifetimes of both are equally reduced by oxygen—but its properties are distinctly different from those of **3**³. The triplet energy of **3** (*E*_T = 70.5 kcal mol⁻¹,^{2c} DFT calculation;⁷ *E*_T = 69.5 kcal mol⁻¹) is well above that of naphthalene. On the other hand, the excitation energy of the quinonoid enol **3**[′] is expected to be much lower

(5) Naphthalene concentrations ≤ 1 mM, selective excitation of the ketone at 351 nm (XeF excimer laser). The formation of triplet naphthalene was monitored at λ_{max} = 415 nm.

(6) Spectrophotometric titration in tris(hydroxymethyl)amine/HCl buffer (ionic strength *I* = 0.1 M). See also: Vandenberg, J. M.; Henrich, C.; Vandenberg, S. G. *Anal. Chem.* **1954**, *26*, 726–727.

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(8) Bromophenol blue is protonated by **3**³ at a diffusion-controlled rate (λ_{obs} = 600 nm), but the proton transfer is largely suppressed when **3**³ is quenched by oxygen (1 atm).

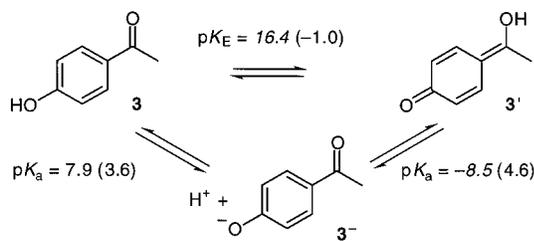
[‡] University of Kansas.

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(4) Excimer laser operated on XeCl, 308 nm, ~100 mJ per pulse, pulse width 25 ns. Low concentrations of ketones **1**, **3**, and **4** were used to avoid radical formation by intermolecular hydrogen abstraction: Das, P. K.; Encinas, M. V.; Scaiano, J. C. *J. Am. Chem. Soc.* **1981**, *103*, 4154–4162. Canonica, S.; Hellrung, B.; Wirz, J. *J. Phys. Chem. A* **2000**, *104*, 1226–1232.

Scheme 1. Ionization and Tautomerization Equilibria of **3**^a

^aValues for the lowest triplet states are given in brackets. Values relying on DFT calculations⁷ are in italic.

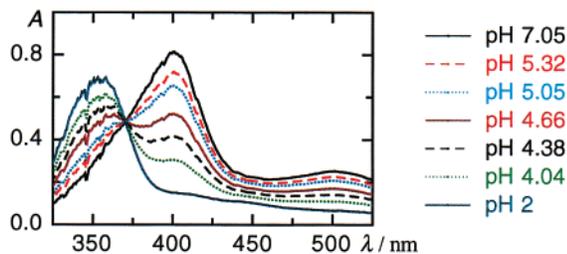


Figure 1. Transient absorption spectra obtained by LFP of **3** in water (10% AN) with various buffers.

(DFT calculation:⁷ $E_T = 38.4$ kcal mol⁻¹). Indeed, **33'** fails to sensitize naphthalene,⁵ $k_{et} < 1 \times 10^8$ M⁻¹ s⁻¹.

DFT calculations for the lowest triplet state correctly predict that enol **33'** is favored over phenol **33**, $pK_E^* = -7.0$, but the calculated value is too low. The triplet energy of **33'** is $E_T(\mathbf{33}') = 64.7$ kcal mol⁻¹.⁹ Ignoring entropy terms, the enolization constant of **33** may be calculated as $pK_E^* = pK_a(\mathbf{33}) - pK_a^*(\mathbf{33}') + \{E_T(\mathbf{33}') - E_T(\mathbf{33})\}/(2.3RT) = 7.9 - 4.6 + \{64.7 - 70.5\}/1.36 \approx -1.0$, and the acidity constant of **33** as $pK_a^*(\mathbf{33}) = pK_E^* + pK_a^*(\mathbf{33}') \approx 3.6$ (Scheme 1). This is consistent with the observed rate of ionization, $k_{ion} \approx 9 \times 10^6$ s⁻¹ in aqueous AN (50% H₂O): $K_a^*(\mathbf{33}) = k_{ion}/k_{H^+}$. The observed fast ionization of **33**, followed by diffusional re-protonation of the resulting anion **33-** to form enol **33'**, excludes a concerted, "intramolecular"^{3a} proton-transfer mechanism through water.

Yet another intermediate, $\lambda_{max} \approx 325$ nm, is formed by LFP of **3** in neutral aqueous solutions. The yield and the lifetime of this transient are not affected by oxygen, and its absorption spectrum is identical with that of the anion **33-** in aqueous base, as noted by Wan and co-workers.³ The main precursor of **33-** is the triplet anion **33-**: a resolved absorbance growth of **33-** at 325 nm, $k = 5 \times 10^6$ s⁻¹, matches the decay of **33-** that is observed at $\lambda \geq 400$ nm in oxygen-saturated water. The subsequent decay of **33-** is not first order in the absence of buffers, because pH increases as **33-** is protonated to **33**.

LFP⁴ of diethyl phosphate **1** in carefully dried, degassed AN showed a strong transient absorbance, $\lambda_{max} = 395$ nm, which is assigned to **31**. The decay rate of **31** increased linearly with increasing concentration of **1** (0.1–1.5 mM, self-quenching rate constant $k_{sq} = 8.5 \times 10^8$ M⁻¹ s⁻¹, intercept $k_0 = 2 \times 10^5$ s⁻¹). Self-quenching is attributed to head-to-tail hydrogen abstraction,⁴ which leads to the radical-derived photoproducts found in dry AN. Energy transfer to naphthalene,⁵ $k_{et} = 7.8 \times 10^9$ M⁻¹ s⁻¹, identified **31** as an excited triplet. The decay rate of **31** increased upon admission of air or oxygen, $k_q \approx 3 \times 10^9$ M⁻¹ s⁻¹, and upon addition of piperylene (0–1.25 $\times 10^{-2}$ M), $k_q = 4.5 \times 10^9$ M⁻¹ s⁻¹. The rate constant increased in a parabolic fashion with

(9) Phosphorescence of **33-** was measured at 77 K in glassy ethanol over solid Na₂CO₃, $\lambda_{max}(0-0) = 442$ nm, $\tau_{ph} = 0.79$ s.

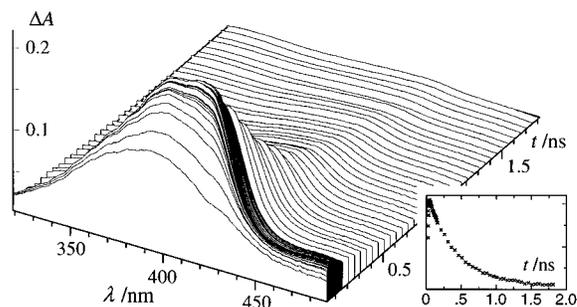


Figure 2. Pump-probe spectra of **1** in aqueous AN (1:1). The kinetic trace shows the temporal evolution of the loading coefficients of the first eigenspectrum obtained by factor analysis.

increasing water content, reaching 2.8×10^7 s⁻¹ at 5% of water, and the transient escaped detection by nanosecond LFP at water contents exceeding 10%.

Excellent agreement between the effect of piperylene on the steady-state and time-resolved data establishes that the triplet state is the reactive excited state in the photorelease of diethyl phosphate from **1**: the ratio of the quantum yields obtained with (10 mM) and without piperylene in 5% aqueous AN, $\phi^q/\phi^0 = 0.39$ (vide supra), is equal, within the limits of error, to the corresponding ratio of the lifetimes of the transient intermediate **31**, $\tau^q/\tau^0 = k_{obs}/(k_{obs} + k_q[q]) = 0.38$.

Addition of concentrated HClO₄ (1–100 $\times 10^{-4}$ M) to AN resulted in a titration plot similar to that shown in Figure 1. Half-protonation was reached at about 4×10^{-3} M acid. The acidic form of the transient is attributed to the quinonoid enol triplet **31'**. Zhang et al. noted that the quantum yield of the photoreaction of *p*HP acetate drops in aqueous acid.^{3a} We observe a similar, if less pronounced drop in acidic aqueous (5%) AN, ϕ ([HClO₄]/M): 0.94 (1 $\times 10^{-4}$), 0.90 (0.05), 0.84 (0.2), 0.74 (1.0). This drop is attributed to protonation of **31-** to the neutral enol **31'**, which is less prone to the release of diethyl phosphate. We could not perform quantum yield measurements in aqueous base, because **1** is quickly hydrolyzed in the ground state. The drop observed at high pH with *p*HP esters³ may be due to a low quantum yield of ISC. We found that excitation of **33-** gives low yields of **33'**, especially at excitation wavelengths of less than 350 nm.

The sharp increase of the decay rate of **31** with increasing water content in AN is attributed to acceleration of heterolytic phosphate release by water. The spectral evolution seen by pump-probe spectroscopy of **1** in aqueous AN (1:1) is shown in Figure 2. Factor analysis of 79 spectra recorded with time delays of 5–2000 ps required 2 spectral components. Adequate fit to the loading factors was obtained with a dual exponential function, $k_1 = (3.8 \pm 0.2) \times 10^{11}$ s⁻¹ (**1** → **31**) and $k_2 = (2.4 \pm 0.2) \times 10^9$ s⁻¹ (phosphate release). We believe that phosphate release occurs directly from the neutral **31**, but rate-determining ionization to **31-** followed by rapid release cannot be excluded.

In summary, the photorelease of diethyl phosphate from *p*-hydroxyphenacyl diethyl phosphate (**1**) in neutral aqueous media proceeds via the triplet state, **31**, which is formed within a few ps and has a lifetime of 0.4 ns in 1:1 aqueous acetonitrile. The subsequent steps of the rearrangement and hydrolysis to *p*-hydroxyphenyl acetic acid (**2**) remain to be elucidated.

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